

Cooperative Self-Assembly of Gold Nanoparticles on the Hydrophobic Surface of Vesicles in Water

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Supporting Information

ABSTRACT: Adsorption of gold nanoparticles (NPs) on a hydrophobic fullerene bilayer vesicle ca. 30 nm in diameter occurs through cooperation of vesicle/NP and NP/NP interactions to produce a NP-vesicle hybrid whose surface is uniformly covered with the NPs separated from each other by a few nm. The vesicle coverage by NPs makes the NP-vesicle hybrid unusually stable to withstand high temperature, chromatographic purification, and high salt concentration-conditions too harsh for ordinary selfassembled vesicles, such as lipid vesicles, to survive. The hybrid serves as a platform of chemical reactions; for example, gold-catalyzed reduction of an aromatic nitro group and deposition of gold atoms for in situ growth of the NPs from 3.5 to 7.2 nm in diameter. The robust vesicle structure can be destroyed by the heat produced in interparticle plasmon coupling absorption of a 532 nm laser irradiation.

Mesoscopic assemblies of plasmonic nanoparticles (NPs) show collective properties that are produced via coupling of the surface plasmon resonance of neighboring NPs.^{1,2} The assembly of NPs on a lipid vesicle, therefore, has been examined as a tool for vesicle functionalization for biological and material applications such as light-triggered release of contents and cell imaging.³⁻⁶ A number of problems have, however, been found as to the stability and control of the density and distribution of NPs on the vesicle, and, most fundamentally, fusion and aggregation of lipid vesicles upon decoration with NPs.^{7–10} Here, we report the circumvention of these problems by the use of a bilayer vesicle (V1-V4) made of a fullerene amphiphile R_5C_{60} ^{-K+} (1-4, in the order of decreasing hydrophobicity; Figure 1),¹¹⁻¹³ which is decorated with Au NPs coated with (11-mercaptoundecyl)tetra(ethylene glycol) groups (NP-1). These oligo(ethylene glycol) (OEG) chains are capable of binding to the potassium countercation of the anionic vesicle. The decoration of V1 with NP-1 (3.5 ± 1.0 nm) took place selectively on one vesicle after another to produce a fully decorated vesicle/NP hybrid (Figure 1a) that undergoes neither aggregation nor fusion. The vesicle, decorated densely with a monolayer of NP-1 (hereafter hybrid), is stable enough to withstand chromatography purification, and to allow in situ growth of the NPs to 7.2 nm upon treatment with HAuCl₄/formaldehyde (Figure 1a). The plasmon effects for the 7.2 nm diameter NP on V1 were detected spectroscopically and photothermally under laser



Figure 1. Self-assembly and growth of NP-1 on fullerene vesicles V1– V4. (a) Overall scheme. (b) Binding between the V1 and NP-1.

irradiation. We consider that a vesicle/potassium-bound NP attractive interaction between the negative V1 surface and the positive NP-1/potassium complex (Figure 1b) and the NP/NP attractive interaction operates cooperatively to allow complete NP-coating of V1. It is probably this cooperation that is responsible for the stability of the V1/NP-1 hybrid (vide infra).

Water-soluble NP-1^{14–16} (3.5 \pm 1.0 nm in diameter of the Au core determined by scanning transmission electron microscopy, STEM. See SI) was synthesized using the Brust method.^{17,18} The hydrodynamic diameter determined by dynamic light scattering (DLS) was 8.2 ± 0.1 nm. NP-1 (concentration of gold atoms = 3.6 mM, 25 °C) shows airwater interface activity,^{19,20} as demonstrated by the decrease in the surface tension of a water droplet in air (72.0 mN m⁻¹) to 46.8 mN m⁻¹ (Figure 2a and b left, gray line; fitting to the Young-Laplace equation). Fullerene vesicles V1-V4 with an average hydrodynamic diameter of 27-47 nm were prepared in water by spontaneous self-assembly of fullerene amphiphiles 1-4, respectively, as we previously reported.^{11,12} We then coated the vesicle surface with the NPs. The vesicles V1-V4 expose hydrophobic alkyl or perfluoroalkyl group at the aqueous interface.

Mixing V1 (hydrodynamic diameter of 30.4 ± 0.5 nm in pure water) and NP-1 resulted in spontaneous formation of the hybrid (Figure 1a; Figure 2e-g for images by STEM and scanning electron microscopy, SEM), which is stable enough to be separated from free NPs by Sephacryl (dextran/bis-

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Figure 2. Adsorption of NP-1 on V1–V4. (a) Reduction of the air/ water surface tension caused by the release of NPs as measured by water droplet pendant method. (b) Time-dependent change of the surface tension of a water droplet in air containing NP-1-decorated V1–V4 (cf. Figure 1a). Experiments using a NP/vesicle ratio of 11 are shown on the left graph, and a ratio of 22 on the right. The vesicles have no surface activity under the experimental conditions (<1 mM).²¹ Further experimental details are given in the SI. (c) Separation of unbound NPs (top band) from decorated vesicles (bottom band) through a Sephacryl (dextran/bis(acrylamide)) size-exclusion column. (d) DLS size distribution of V1 before (black line) and after (red line) NP decoration. (e) STEM image of decorated vesicles. The average number of NPs on a vesicle is 22.6 ± 1.7 NP/vesicle. Scale bar is 100 nm. (f, g) STEM and SEM images of a decorated vesicle. Scale bar is 10 nm.

(acrylamide)) size-exclusion chromatography (Figure 2c). The hydrodynamic diameter of V1 increased from 30.4 ± 0.5 to 49.4 \pm 0.2 nm (Figure 2d). The ζ -potential of the vesicles, $-45.6 \pm 2.4 \text{ mV} \text{ (pH} = 7.0\text{), became } -13.7 \pm 0.8 \text{ mV; a value}$ similar to that of NP-1 (-14.9 \pm 3.1 mV). The hydrodynamic diameter of the NP/vesicle hybrid did not change upon heating at 80 °C, providing an additional evidence of this stability. Whereas V1 immediately precipitates at high salt concentration, the NP-1-V1 hybrid remains stable in phosphate-buffered saline for 8 h at room temperature with ca. 10% increase in the diameter, and the size increase gradually slows down with time (SI). This is probably because NP-decoration converted the ionic vesicle surface into a neutral ethylene glycol-covered surface (Figure 1b). The gold NPs can be recovered from the hybrid by protonation of the vesicle followed by removal of the resulting neutral fullerene by extraction with an organic solvent.

Surface tension measurements of a mixture of NP-1 and vesicles V1-V4 at the air-water interface indicated particularly strong affinity of NP-1 to V1. Thus, we prepared a droplet from the mixture (time = 0) and measured the time-dependent change of surface tension caused by the movement of NP-1 from the vesicle surface to the air-water interface (Figure 2a). The surface tension of a reference solution of NP-1 ([Au] =0.36 mM) is 46.8 \pm 0.4 mN m⁻¹. The first set of experiments was performed for a NP-1/vesicle ratio of ca. 11. The surface tension of the solution $(70.7-71.6 \text{ mN m}^{-1})$ gradually decreased as NP-1 moved to the air-water interface (Figure 2b, left). For V1, the surface tension decreased slightly after 15 min to 70.3 \pm 0.3 mN m⁻¹ and 69.0 \pm 1.9 for V2 and significantly to 58.6 \pm 0.3 for V3 and 50.1 \pm 2.8 for V4, indicating the binding of NP-1 to V1-V4 decreases as the hydrophobicity of the vesicle surface decreases. Further experiments for V1 and V2 with a ratio of 22 indicated that NP-1 hardly dissociates once adsorbed on V1 (Figure 2b, right). Thus, we conclude that one of the driving forces for NP-1 binding to V1 is the hydrophobicity of the OEG chains when complexed with potassium cation (cf. ability of crown ether to bind on a potassium cation in a hydrophobic solvent).^{10,22} The stability of the hybrid at the high salt concentration (vide supra) supports the mechanism. Therefore, we focused on this combination.

The NP-1–V1 hybrid is stable even under electron irradiation in vacuum under the STEM conditions (Figure 2e–g). The STEM vesicle size increased from 30.8 ± 5.2 to 37.7 ± 4.0 nm. The difference of 6.9 nm is twice the size of the gold core of NP-1. All vesicles are fully covered with NP-1 with high uniformity of distribution on the surface (STEM, Figure 2e). The NPs are separated from each other by ca. 2 nm (Figure 2f), which is much shorter than the stretched OEG side chain (2.9 nm) and suggests that in each vesicle, OEG chains interact with each other in the void between the Au cores.^{2,23,24}

Experiments using a deficient amount of NP-1 provided evidence for the cooperation of the two forces in the vesicle/ NP-1 hybrid formation: the vesicle/potassium-bound NP electrostatic attraction (Figure 1b) and the potassium-bound NP/NP hydrophobic interaction. Here, we found that most of the vesicles are either fully or lightly coated by NPs (STEM, Figure 3a and I and II). This observation is quantified for the correlation shown in Figure 3b blue bars (for [Au]/[1] = 2.25; i.e., deficiency of NPs), which indicates that the vesicles are either lightly coated (0 < n < 7) or densely coated (13 < n < 33; note that the number *n* is related to the diameter of the vesicle). Figure 3b (red bars) provides an additional illustration that a large number of NPs are located on densely decorated vesicles.

The number of vesicles bearing NPs decreases for n = 1-7 and then increases above n = 13-ca. 25 (blue). In Figure 3c (blue dots) we see a linear correlation between the number of NP-coated vesicles and the [Au]/[1] ratio, while in Figure 3c (red dots) we see that the average number of NP-1 on each vesicle is nearly independent of the [Au]/[1] ratio. These data agree with a cooperative assembly mechanism initiated by reversible formation of small nuclei followed by rapid irreversible growth of assemblies²⁵ on the two-dimensional surface of the vesicle.²⁶

Interestingly, the mechanism of assembly of Au NPs on V1 depends on the structure of the OEG side chain. When we used a NP coated with longer chains, (11-mercaptoundecyl)hexa-(ethylene glycol) (NP-2, 3.2 ± 1.5 nm in diameter of the Au core by STEM and hydrodynamic diameter of 13.2 ± 0.1 nm



Figure 3. Adsorption of NP-1 and NP-2 on vesicle V1. (a) STEM image of cooperative adsorption of NP-1 on V1. Scale bar is 100 nm. (b) The number of vesicles bearing an *n* number of NP-1/V1 examined for 339 vesicles. The blue bars indicate the number of vesicles (*x*-axis) bearing *y*-number of NPs on a vesicle. The red bars indicate the total number of NPs on vesicles having *y*-number of NPs. (c) The ratio of NP-1 vs V1 (shown in [Au] and [1] employed for the experiments). The red bars indicate the total number of NPs. (d) STEM image of isodesmic decoration of NP-2 on V1 that results in vesicle agglomeration and fusion. Inset shows a large vesicle because of fusion. Scale bars are 100 nm. (e) The [Au]/[1] ratio affecting hydrodynamic diameter of assemblies of the hybrid for NP-1 and NP-2. Determined by DLS at 25 °C.

by DLS), the adsorption process became isodesmic probably because of the decreased hydrophobic interaction among potassium-bound NP-2 (i.e., larger water solubility of NP-2 than NP-1), and the NP-decoration became controlled largely by the vesicle-NP interaction (Figure 1b). Partial decoration of V1 with NPs resulted in neutralization of the negative charge of the vesicle surface (increased ζ -potential) and hence promoted vesicle agglomeration and fusion, as seen in Figure 3d and inset, respectively.²⁷ The DLS data in Figure 3e also illustrate the huge difference of the distribution of the size of vesicle/NP hybrids between the NP-1 and NP-2 hybrids. A similar phenomenon has been reported for lipid vesicles, where NP causes deformation of the bilayer membrane and vesicle fusion caused by charge neutralization.⁸

NP-1 on V1 serves as a platform for chemical reactions. NP-1–V1 hybrid (ca. 11 NPs/vesicle) shows a catalytic activity in the NaBH₄ reduction of 4-nitrophenol (SI), while neither NP-1 in water without vesicle nor V1 show any catalytic activity, indicating the presence of a small area of ligand-free surface on NP-1.²⁸ Another example is the in situ deposition of gold atoms on the adsorbed NP-1 without destruction of the vesicle structure (Figure 4a–c). Thus, the diameter of NP-1 was



Figure 4. In situ growth of Au NPs on the surface of V1 and light-triggered collapse. (a) STEM and SEM images of the hybrid after growth of Au NPs ($[HAuCl_4]/[1] = 150$). Scale bar is 100 nm. Insets show magnified STEM (top) and SEM (bottom) images of the hybrid (scale bar 10 nm). (b) The ratio of HAuCl₄ vs V1 (shown in [1] used) affecting the diameter of the Au NP and the absorption peak wavelength of the surface plasmon resonance. (c) Absorption spectra of free NPs and NPs before and after destruction of the vesicle structure. (d) SEM (I) and STEM images (II) of collapsed vesicles after laser irradiation of V1 bearing 7.2 nm NPs for 30 min. Loss of the NP and vesicle images is seen. Scale bar is 100 nm.

increased from 3.5 ± 1.0 to 7.2 ± 1.4 nm by adding a solution of HAuCl₄/K₂CO₃ (1:3) and formaldehyde (excess) to a solution of **NP-1** on **V1** (2 h at room temperature). By changing the amount of the added gold source ([HAuCl₄]/[1] in Figure 4b), we noted that the growth stops at 7.2 ± 1.4 nm (black circles), probably because further growth disturbs the OEG network or the growth of a ligand-free surface on the Au NP counterbalances the NP growth. Note that attempts failed to directly decorate **V1** with large Au NPs (6.7 nm in diameter). The observed contact inhibition makes an interesting contrast to the growth of bare Au NPs under the same conditions to a size of 40–50 nm on a polymer-coated carbon nanotube substrate.²⁹

The diameter increase accompanied a shift in the NP absorption from 510 to 544 nm (Figure 4b, red squares), which started when the NP grew larger than 4.6 ± 1.1 nm, suggesting the occurrence of plasmonic resonance coupling at this size. The absorption spectra of the NP-1–V1 hybrid before and after growth are shown in Figure 4c and compared with the spectrum of NP-1 collected after protonative destruction of V1 (517 nm, as expected for a 7 nm Au NP).³⁰ Laser irradiation (532 nm, 20 mW, 30 min) of the postgrown NPs of 7.2 nm diameter³¹ caused heating by plasmonic effects³² and destroyed both the NPs and the vesicle (formation of a featureless object in Figure 4d). Laser irradiation of the hybrid bearing the low-

melting point 3.4 nm NPs resulted in the loss of the NP images and the formation of large Au particles, suggesting that the NPs melted away.

In conclusion, we have decorated a hydrophobic vesicle V1 through interfacial adsorption of OEG-coated gold NPs in water. By selecting NP-1 instead of NP-2 that has longer OEG chains, we decorated one vesicle after another with the NPs because of cooperation of two binding forces, electrostatic attraction between the anionic vesicle and the cationic NP/ potassium complex, and hydrophobic interaction among potassium-bound NPs. Here a standard two-step mechanism of nucleation/growth mechanism is expected to occur.³³ On the other hand, NP-2 randomly decorates many vesicles, partially neutralizes the negative charge of the vesicle, and causes vesicle agglomeration and fusion. The data on NP-1 thus represent a rare case of cooperative assembly on a spherical surface.²⁶ The fullerene vesicle is robust enough to allow sizecontrolled metal atom deposition, suggesting applications for the preparation of multimetallic core-shell structures.

ASSOCIATED CONTENT

Supporting Information

Experimental procedures, methods, additional electron microscopy images, and absorption spectra. The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/jacs.5b03632.

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Notes

The authors declare no competing financial interest.

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REFERENCES

(1) Song, J.; Zhou, J.; Duan, H. J. Am. Chem. Soc. 2012, 134, 13458–13469.

- (2) Niikura, K.; Iyo, N.; Higushi, T.; Nishio, T.; Jinnai, H.; Fujitani, N.; Ijiro, K. J. Am. Chem. Soc. **2012**, 134, 7632–7635.
- (3) Skirtach, A. G.; Dejugnat, C.; Braun, D.; Susha, A. S.; Rogach, A. L.; Parak, W. J.; Möhwald, H.; Sukhorukov, G. B. *Nano Lett.* **2005**, *5*, 1371–1377.
- (4) Jain, P. K.; Huang, X.; El-Sayed, I. H.; El-Sayed, M. A. Acc. Chem. Res. 2008, 41, 1578-1586.
- (5) Wu, G.; Mikhailovsky, A.; Khant, H. A.; Fu, C.; Chiu, W.; Zasadzinski, J. A. J. Am. Chem. Soc. 2008, 130, 8175–8177.
- (6) Mart, R. J.; Liem, K. P.; Webb, S. J. Pharm. Res. 2009, 26, 1701–1710.
- (7) Volodkin, D. V.; Skirtach, A. G.; Möhwald, H. Angew. Chem., Int. Ed. 2009, 48, 1807–1809.
- (8) Michel, R.; Gradzielski, M. Int. J. Mol. Sci. 2012, 13, 11610–11642.
- (9) Zhang, L.; Granick, S. Nano Lett. 2006, 6, 694-698.
- (10) Van Lehn, R. C.; Ricci, M.; Silva, P. H. J.; Andreozzi, P.; Reguera, J.; Voïtchovsky, K.; Stellacci, F.; Alexander-Katz, A. Nat. Commun. 2014, 5, 4482.

(11) Homma, T.; Harano, K.; Isobe, H.; Nakamura, E. Angew. Chem., Int. Ed. 2010, 49, 1665.

- (12) Homma, T.; Harano, K.; Isobe, H.; Nakamura, E. J. Am. Chem. Soc. 2011, 133, 6364–6370.
- (13) Burger, C.; Hao, J.; Ying, Q.; Isobe, H.; Sawamura, M.; Nakamura, E.; Chu, B. J. Colloid Interface Sci. 2004, 275, 632–641.
- (14) Glogowski, E.; Tangirala, R.; He, J.; Russel, T. P.; Emrick, T. Nano Lett. 2007, 7, 389–393.
- (15) Simpson, C. A.; Huffman, B. J.; Gerdon, A. R.; Cliffel, D. E. Chem. Res. Toxicol. 2010, 23, 1608-1616.
- (16) Alkilany, A. M.; Murphy, C. J. J. Nanopart. Res. 2010, 12, 2131–2333.
- (17) Brust, M.; Bethell, D.; Schiffrin, D. J.; Kiely, C. J. Adv. Mater. 1995, 7, 795–797.

(18) Kanaras, A. G.; Kamounah, F. S.; Schaumburg, K.; Kiely, C. J.; Brust, M. Chem. Commun. **2002**, 2294–2295.

- (19) Isa, L.; Calzolari, D. C. E.; Pontoni, D.; Gillich, T.; Nelson, A.; Zirbs, R.; Sánchez-Ferrer, A.; Mezzenga, R.; Reimhult, E. *Soft Matter* **2013**, *9*, 3789–3797.
- (20) Binder, W. H. Angew. Chem., Int. Ed. 2005, 44, 5172-5175.
- (21) Harano, K.; Yamada, J.; Mizuno, S.; Nakamura, E. *Chem.–Asian J.* **2015**, *10*, 172–176.
- (22) Tracy, J. B.; Kalyuzhny, G.; Crowe, M. C.; Balasubramanian, R.; Choi, J.-P.; Murray, R. W. J. Am. Chem. Soc. **2007**, 129, 6706–6707.
- (23) DeVries, G. A.; Brunnbauer, M.; Hu, Y.; Jackson, A. M.; Long, B.; Neltner, B. T.; Uzun, O.; Wunsch, B. H.; Stellacci, F. *Science* **2007**, *315*, 358–361.
- (24) Zheng, Y.; Lalander, C. H.; Bach, U. Chem. Commun. 2010, 46, 7963–7965.
- (25) Harano, K.; Homma, T.; Niimi, Y.; Koshino, M.; Suenaga, K.; Leibler, L.; Nakamura, E. *Nat. Mater.* **2012**, *11*, 877–881.
- (26) Zhang, K.-Q.; Liu, X. Y. Nature 2004, 429, 739-743.
- (27) Harano, K.; Narita, A.; Nakamura, E. Chem. Lett. 2014, 43, 877–879.

(28) (a) Biswas, M.; Dinda, E.; Rashid, Md. H.; Mandal, T. K. J. Colloid Interface Sci. 2012, 368, 77–85. (b) Lee, H.-Y.; Shin, S. H. R.; Abezgauz, L. L.; Lewis, S. A.; Chirsan, A. M.; Danino, D. D.; Bishop, K.

J. M. J. Am. Chem. Soc. **2013**, 135, 5950–5953.

(29) Wang, X.; Wang, C.; Cheng, L.; Lee, S.-T.; Liu, Z. J. Am. Chem. Soc. 2012, 134, 7414-7422.

(30) Nguyen, N. D.; Dang, V. P.; Le, A. Q.; Nguyen, Q. H. Adv. Nat. Sci. 2014, 5, 045002.

- (31) Castro, T.; Reifenberger, R.; Choi, E.; Andres, R. P. Phys. Rev. B 1990, 13, 8548-8556.
- (32) Govorov, A. O.; Richardson, H. H. Nano Today 2007, 2, 30-38.
- (33) Vekilov, P. G. Nat. Mater. 2012, 11, 838-840.